

# From plants to products O blanhigion i gynhyrchion

Dr. Sreenivas Rao Ravella Senior fermentation scientist rsr@aber.ac.uk









#### **IBERS**

#### Institute of Biological, Environmental and Rural Sciences



#### IBERS - staff

**Plant breeders Agronomists Molecular geneticists Biochemists Physiologists** Rumen/silage expertise **Enzymes/microbiologists** Other sources

#### **Phenomics Centre**



# Platform for non-destructive dynamic imaging of plant growth & development



Controlled environment conveyor based system radio-tagged plants



# state-of-the art imaging stations:

- visible
- near IR
- Thermal IR
- fluorescence
- laser scan 3-d
- both canopy and root imaging
- DNA sequencing for rapid phenotypic associations

#### **BEACON** and what it will do

- Welsh European Funding Office (WEFO) funded initiative with a value of £20m
- Partnership between Aberystwyth, Swansea and Bangor Universities to develop biorefining R&D expertise in Wales
- Enable academic and a wide range of industrial partners to develop and demonstrate scale-up processes for economically viable industrial applications

### **WEFO Metricated Objectives**

#### **Outputs**

- 202 enterprise assists
- 25 R & D collaborations

Enterprise has to be in Wales (or have a registered office)

If leading to inward investment then outside companies can also be counted

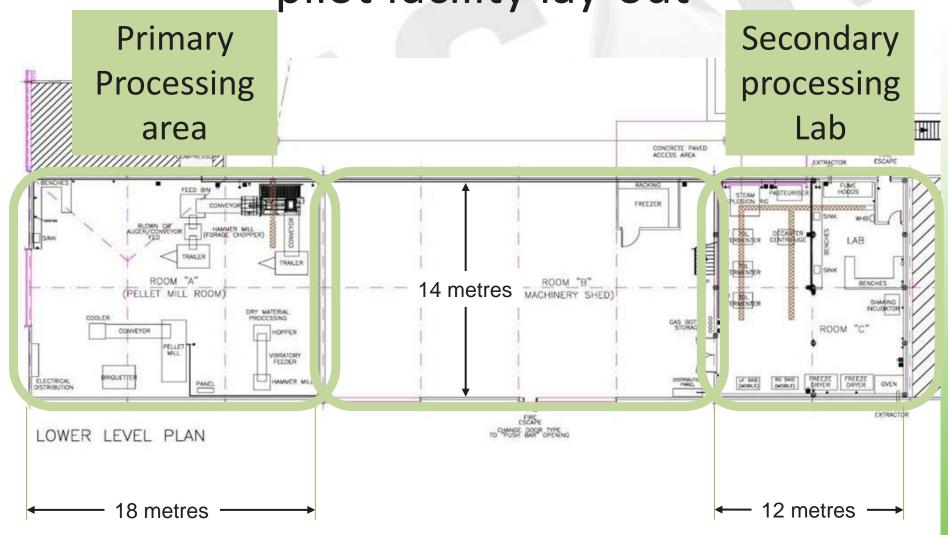
#### Results

- 67 Jobs Created
- 3 Enterprises Created
- Profit Benefit £1,680,000
- Investment Induced £3,360,000
- 7 Products, process or services registered
- 16 New/improved products, processes or services launched

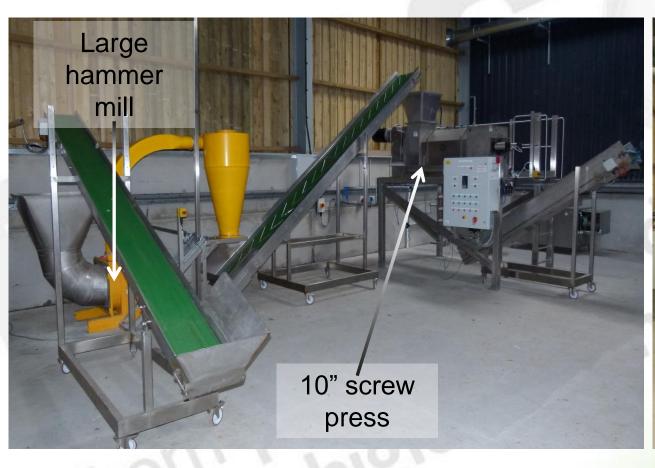
### **BEACON**

Facilities and equipment for development of biorefining expertise and process scale-up

Biomass treatment and fermentation pilot facility lay-out



### **Primary Processing Area**





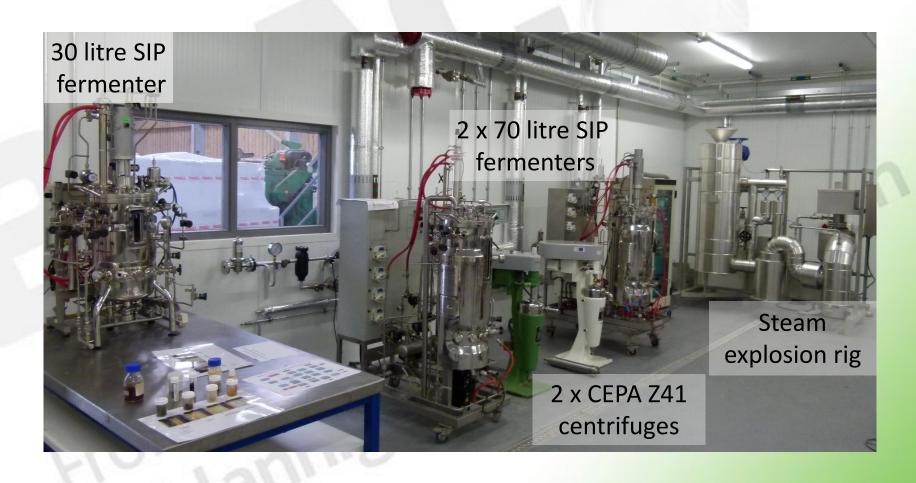
Integrated wet processing line

Dry processing: Pellet mill

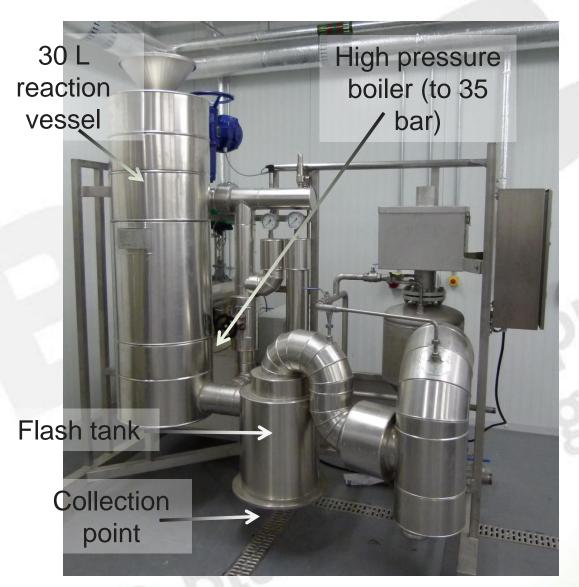




# Secondary processing Lab Fermentation and pre-treatment



### Steam explosion rig by Cambi



Pre-treated biomass from the flash tank



SE is common scalable pretreatment for lignocellulosic biomass. Above material will liquefy in under 1 hour with standard cellulosic enzymes.











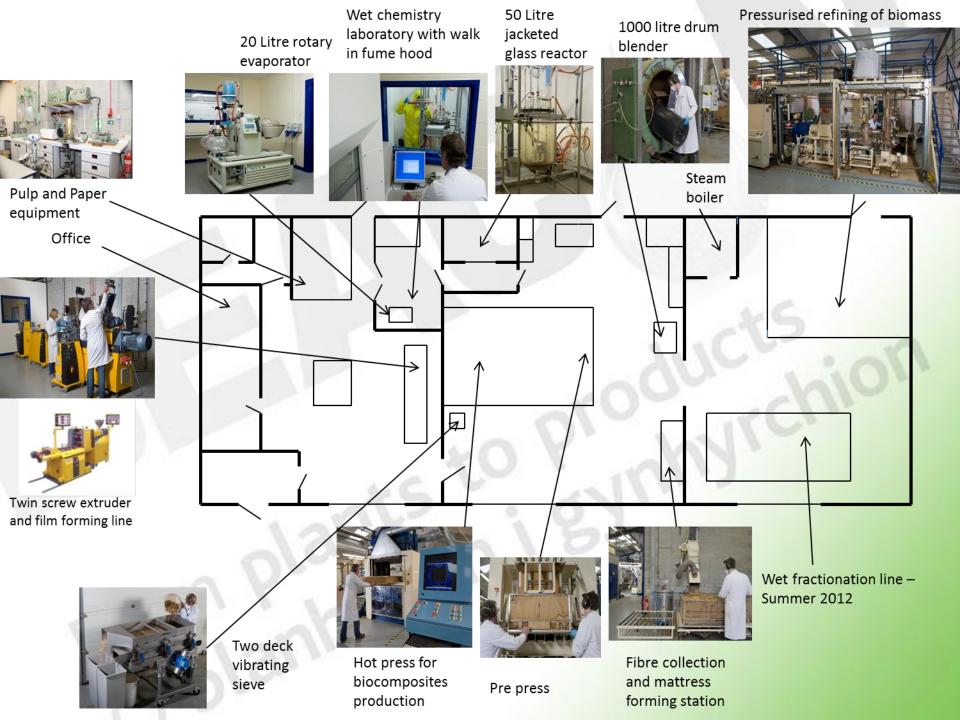


### **Biochar Facility**



Hot gas recirculation

Condenser for pyrolysis oils



#### Climate-KIC







Climate-KIC

For Students For Entrepreneurs For Businesses For Public Bodies bout Us

Climate-KIC

Climate-KIC > NL > Innovation and Pathfinder projects 2012

Innovation and Pathfinder projects 2012

Final decisions about the Innovation and Pathfinder Projects for 2012 are made; Dutch partners are well represented.

Tuesday 25th October

Our Next Event

View all



31 May 2013, Netherlands

Seminar: Innovative financial arrangements

Climate-KIC News View all



22nd May 2013

Climata I/IC antronyonous wir

News

Case Studies

Events

Videos

# Samples from university of Stirling

- aquatic plant material
- composted Elodea, pondweed (Lemna sp) and Water Lily







# Enzymatic hydrolysis



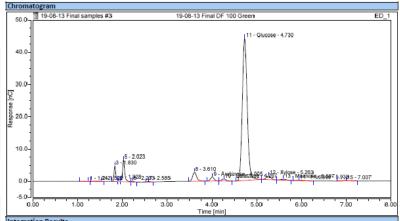






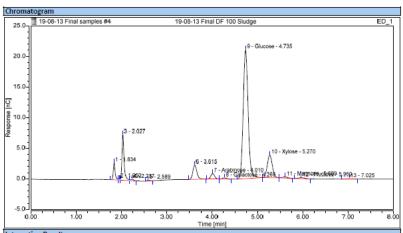
## Hydrolysis results

#### **Chromatogram and Results** Injection Details 19-08-13 Final DF 100 Green Run Time (min): 8.00 Injection Name: Vial Number: Injection Volume: 10.00 Injection Type: Unknown Channel: ED\_1 Calibration Level: Wavelength: n.a. Instrument Method: Basic mono method Bandwidth: Processing Method: New ProcMethod Dilution Factor: 1.0000 Injection Date/Time: 19/Aug/13 12:32 Sample Weight: 1.0000



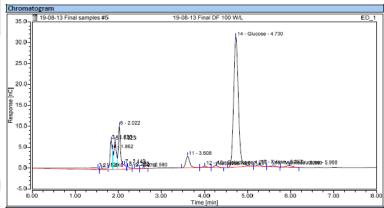
Integ	gration Results						
No.	Peak Name	Retention Time	Area	Height	Relative Area	Relative Height	Amount
		min	nC*min	nC	%	96	ug/ml
1		1.242	0.007	0.079	0.10	0.13	n.a.
2		1.522	0.016	0.075	0.23	0.12	n.a.
3		1.830	0.220	4.826	3.15	7.65	n.a.
4		1.928	0.026	0.510	0.37	0.81	n.a.
5		2.023	0.389	6.794	5.56	10.77	n.a.
6		2.233	0.016	0.239	0.23	0.38	n.a.
7		2.585	0.014	0.203	0.21	0.32	n.a.
8		3.610	0.291	2.792	4.16	4.43	n.a.
9	Arabinose	4.005	0.115	1.211	1.65	1.92	0.2339
10	Galactose	4.267	0.068	0.636	0.97	1.01	0.1385
11	Glucose	4.730	5.615	44.112	80.26	69.95	11.4813
12	Xylose	5.263	0.140	1.113	2.00	1.76	0.2987
13	Mannose	5.597	0.037	0.253	0.52	0.40	0.0810
14	Fructose	5.938	0.019	0.111	0.28	0.18	0.0677
15		7.007	0.022	0.109	0.31	0.17	n.a.
Total	:		6.996	63.063	100.00	100.00	

Chromatogram and Results								
Injection Details								
Injection Name:	19-08-13 Final DF 100 Sludge	Run Time (min):	8.00					
Vial Number:	RA4	Injection Volume:	10.00					
Injection Type:	Unknown	Channel:	ED 1					
Calibration Level:		Wavelength:	n.a.					
Instrument Method:	Basic mono method	Bandwidth:	n.a.					
Processing Method:	New ProcMethod	Dilution Factor:	1.0000					
Injection Date/Time:	19/Aug/13 12:41	Sample Weight:	1.0000					



Integ	ration Results						
No.	Peak Name	Retention Time	Area	Height	Relative Area	Relative Height	Amount
		min	nC*min	nC	%	%	ug/ml
1		1.834	0.132	2.773	3.21	7.03	n.a.
2		1.950	0.006	0.170	0.14	0.43	n.a.
3		2.027	0.412	7.461	10.04	18.92	n.a.
4		2.237	0.019	0.237	0.46	0.60	n.a.
5		2.589	0.011	0.172	0.27	0.44	n.a.
6		3.615	0.242	2.333	5.91	5.91	n.a.
7	Arabinose	4.010	0.087	0.898	2.13	2.28	0.1771
8	Galactose	4.269	0.023	0.204	0.55	0.52	0.0460
9	Glucose	4.735	2.639	21.062	64.34	53.40	5.3954
10	Xylose	5.270	0.470	3.689	11.46	9.35	0.9963
11	Mannose	5.609	0.026	0.207	0.63	0.52	0.0573
12	Fructose	5.960	0.028	0.175	0.67	0.44	0.0960
13		7.025	0.007	0.063	0.18	0.16	n.a.
Total:			4.101	39.443	100.00	100.00	

Chromatogram and Results								
njection Details								
Injection Name:	19-08-13 Final DF 100 W/L	Run Time (min):	8.00					
Vial Number:	RA5	Injection Volume:	10.00					
Injection Type:	Unknown	Channel:	ED 1					
Calibration Level:		Wavelength:	n.a.					
Instrument Method:	Basic mono method	Bandwidth:	n.a.					
Processing Method:	New ProcMethod	Dilution Factor:	1.0000					
Injection Date/Time:	19/Aug/13 12:50	Sample Weight:	1.0000					



Integra	ation Results						
No.	Peak Name	Retention Time	Area	Height	Relative Area	Relative Height	Amount
		min	nC*min	nC	%	%	ug/ml
1		1.525	0.352	0.490	5.37	0.74	n.a.
2		1.618	0.061	0.326	0.93	0.49	n.a.
3		1.830	0.360	7.018	5.50	10.54	n.a.
4		1.862	0.104	4.783	1.58	7.18	n.a.
5		1.923	0.434	6.827	6.62	10.25	n.a.
6		2.022	0.702	10.377	10.71	15.58	n.a.
7		2.143	0.010	0.263	0.16	0.39	n.a.
8		2.222	0.044	0.532	0.68	0.80	n.a.
9		2.378	0.015	0.163	0.23	0.24	n.a.
10		2.580	0.014	0.198	0.21	0.30	n.a.
11		3.608	0.287	2.733	4.39	4.10	n.a.
12	Arabinose	4.007	0.031	0.321	0.48	0.48	0.0632
13	Galactose	4.267	0.052	0.486	0.80	0.73	0.1066
14	Glucose	4.730	3.942	31.090	60.20	46.69	8.0597
15	Xylose	5.267	0.041	0.327	0.63	0.49	0.0875
16	Mannose	5.593	0.033	0.237	0.50	0.36	0.0722
17	Fructose	5.968	0.067	0.416	1.02	0.63	0.2321
Total:			6.548	66.587	100.00	100.00	

Dry samples	Sample ID	Mass (g)	%C	%N
Water Lily 1	31	0.1006	42.13	3.975
Water Lily 2 (NaOH)	32	0.0988	34.41	2.6203
Green 1	33	0.0961	39.91	2.3742
Green 2 (NaOH)	34	0.0892	35.67	0.91706
Sludge 1	35	0.0957	26.84	3.6663
Sludge 2 (NaOH)	36	0.09	16.26	0.95165

Wet samples	Sample ID	Mass (g)	%C	%N
Green Fresh 1	37	0.2367	2.115	0.17489
Green Fresh 2	38	0.24	2.091	0.17537
Green Fresh 3	39	0.2047	2.171	0.19118
Water Lily Fresh 1	40	0.2378	4.121	0.33406
Water Lily Fresh 2	41	0.2511	4.56	0.35738
Water Lily Fresh 3	42	0.2199	3.911	0.33194
Liquid Sludge 1	43	0.2406	2.971	0.27358
Liquid Sludge 2	44	0.2404	2.798	0.28151
Liquid Sludge 3	45	0.2634	3.079	0.28861

All samples measured using a C/N elemental analyser At Bangor

					After ashing		1
As received	Sample ID		Vial wt (g)	Fresh wt sample (g)	Vial + sample (g)	ashed sample wt (g)	mg/IP
Green 1		1	13.5564	0.1999	13.5837	0.1726	11.44494
Green 2		2	13.446	0.202	13.4982	0.1498	12.99388
Water Lily 1		3	13.6023	0.1994	13.6249	0.1768	8.519168
Water Lily 2		4	13.4388	0.209	13.5354	0.1124	4.991028
Sludge 1		5	13.4354	0.1953	13.513	0.1177	16.00571
Sludge 2		6	13.6957	0.1535	13.7865	0.0627	19.70595
						1 1 1 ( 1	M. T.

	-d0. 1			After ashing	ALL	
Liquid	Sample ID	Vial wt (g)	Fresh wt sample (g)	Vial + sample (g)	ashed sample wt (g)	mg/IP
Green	7	13.4447	6.8777	13.5662	6.7562	35.53956
Water Lily	8	13.4572	8.5389	13.7665	8.2296	48.36134
Sludge	9	13.4291	6.8319	13.6735	6.5875	74.00489

0.2 g of sample (as received) was ashed at 450oC for 16 hours

or

6.5-9 g of liquid sampe was ashed at 450oC for 16 hours.

1 ml of 20% HCl was then added to the ashed sample, and 9 ml of H2O added.

Concentration in column G is the final concentration of total P (after analyser dilution has been taken account of)

